Gel Extraction / PCR Product Purification Kit (50preps)

Our Gel Extraction /PCR Product DNA fragment isolation kit is used magnetic beads so that is convenient to use and save a money, time, and laboratory space.
The purification procedure removes primers, nucleotides, enzyme, salts, ethidium bromide and other impurities in your DNA containing solutions.

Are you still using column DNA isolation kits, gel extraction kits and PCR clean-up kits? Switch to our kits now to enjoy the high yield recovery.

Switch your Gel extraction kit and PCR clean-up kit to ours and start enjoying the fast purification with magnetic beads.

Advantages and Benefits

- Efficient extraction of DNA fragments from 50-bp to 25-kbp
- High purity of DNA (A260/A280=1.7~1.9)
- Recover DNA fragments from regular or low-melting point agarose gels in TAE or TBE buffer
- Elute DNA with just 15~30 µl elution buffer or ddH_2O
- Recovery up to 98%
- Preparation time: 10~15 minutes
- No shearing of large DNA fragments
- washing buffer-free DNA fragment
- Increase cloning efficiency
- Fast purification process
- No centrifuge needed

DNA purified with the Gel/PCR DNA Isolation System can be used directly in most applications, including:

- Sequencing
- PCR
- Restriction digestion & modifying enzymatic reaction
- DNA ligation for cloning
- Labeling & hybridization

Basically, according to average 100 µg of gel and add 300 µl dissolve buffer.

Reagents	volume
Gel Dissolve buffer (GDB)	15 ml
Magnetic Beads (8 mg/ml)	5 ml
Wash buffer (WB)	45 ml
Elution buffer (EB)	2 ml

Protocol

Cut the DNA band from agarose gel. (Limit size as small as possible). Weight the gel. Dice the gel to small pieces.
Add Gel dissolve buffer (GDB) as 1 (gel weight):3 (volume) (Gel:GDB), for example 100 µg of gel add 300 µl of GDB.
Mix well and set up the tube on heat block for 10 min at 55 °C until gel dissovled in the buffer.
After gel is dissolved, 100 µl magnetic beads is added and shake for 10 second.
Add isopropanol in same volume of the mixture (for example: 100ul gel 300ul GDB 100 beads total 500ul +500 ul isopropanol). Rotate for 2 min at room temperature.
Put the tube on magnetic rack (or magnet) for 30 second (When beads are against the well of tube) and remove the supernatant carefully.
Add 300 µl of wash buffer I and vortex it and put-on magnetic rack for 20 second and remove the supernatant.
Add 300 µl wash buffer II (70% ethanol not included in the kit) and vortex and put it on magnetic rack for 20 second. Remove the supernatant. Repeat this step one more time.
Note: remove wash buffer II completely.
Let the beads dry for 5 min in room temperature. (Do not make it over dry)
Add about 30-50 µl of elution buffer (depend on DNA bands concentration) or ddH_2O. Vortex it and put on the heat block for 3 min at 55°C.
During the time, vortex twice. Put the tube on the magnetic rock and collect DNA fragment.